毛萼香茶菜中的三个新的对映-贝壳杉烷型二萜

王 佳 赵勤实 林中文 孙汉董*

(中国科学院昆明植物研究所植物化学开放实验室,昆明650204)

摘要 从采自云南省江川县的毛萼香茶菜(Isodon eriocalyx)的甲醇提取物中分离得到 3 个新的 6,7-螺断-对映-贝壳杉烷型二萜:表毛萼甲素,毛萼晶 N 和毛萼晶 O;同时还分离到 2 个已知化合物:毛萼甲素及毛萼晶 L,并修正了毛萼甲素的结构。

关键词 唇形科,毛萼香茶菜,二萜

THREE NEW 6,7 -SECO-ENT-KAURANE DITERPENOIDS FROM ISODON ERIOCALYX

Wang Jia, Zhao Qinshi, Lin Zhongwen, Sun Handong*

(Laboratory of Phytochemistry, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204)

Abstract Three new 6,7-seco-ent-kaurane diterpenoids named epi-eriocalyxin A (2), maoecrystal N (3) and maoecrystal O (4) together with two known constituents eriocalyxin A (1) and maoecrystal L (5) were isolated from the dried leaves of *Isodon eriocalyx*. Meanwhile, the structure of eriocalyin A was revised.

Key words Labiatae, Isodon eriocalyx, Diterpenoids

Isodon eriocalyx (Dunn.) Hara, a perennial herb or shrub of Labiatae family, is widely distributed in Southwest China. It has long been used as folk medicine to cure sore throat, inflammation and interdigital disease (吴征镒等, 1977). A series of new ent-kaurane diterpenoids have been isolated from the dried leaves of *I. eriocalyx* collected in five different regions of Yunnan Province, China (Hassan et al, 1994; Takeda et al, 1995).

Further investigation on the same plant led to the isolation and determination of three new 6, 7- seco-ent- kaurane diterpenoids named epi-eriocalyxin A(2), maoecrystal N (3) and maoecrystal O (4) as well as two known constituents eriocalyxin A (1) (王宗玉等, 1982) and maoecrystal L (5) (Shen et al, 1994).

RESULTS AND DISCUSSION

Compound 1, $C_{20}H_{24}O_5$, was obtained as colorless crystals (from MeOH). The spectral data of 1H , ^{13}C NMR, MS, IR, and UV were identical with those of eriocalyxin A reported presviously. The

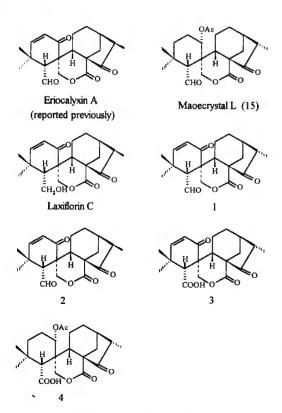
1996-11-07 收稿, 1997-01-10 接受发表

[·]通讯联系人 Author for correspondence

stereochemistry of Me-16 in eriocalyxin A was established as β -orientation, but there was no direct evidence to confirm the determination because of technical limits at that time. Comparison [Table 1] of the ¹³C NMR data of 1 with those of manecrystal L and laxiflorin C (Sun *et al*, 1995), both of which possessed the same skeleton as 1, reminded us to reconsider the orientation of Me-16.

The stereochemistry of Me-16 in laxiflorin C was confirmed as β -orientation by the NOE effects between H-12 β and C-16 methyl, and the unique upfield shift of C-12 (δ 17.9 owing to the γ -effect of Me-16 β . The α -orientation of Me-16 in maoecrystal L was also supported by the significant NOEs between H-14 β and Me-16, and the upfield shift of C-14 (δ 28.4 due to the γ -effect of Me-16 α). Thus, the orientation of Me-16 could be determined by the upfield shift of C-14 or C-12. The upfield shift of C-14 at about δ 28.4 was characterized for Me-16 α while the upfield shift of C-12 at about δ 18 for Me-16 β . Meanwhile, the chemical shift of C-17 was also useful for distinguishing the stereochemistry of Me-16. When the Me-16 was β -orientation, the chemical shift of C-17 was δ 11 or so. Otherwise, the δ value was about δ 17 because of the space effect mentioned above.

In conclusion, the chemical shifts of C-12, 14 and 17 were typical for differentiating the orientation of Me-16.



The chemical shifts of ring C and D in 1 were quite similar to those of maoecrystal L. Consequently, the orientation of Me-16 in 1 should be revised as α. The revised structure of eriocalyxin A was 16(S)-methyl-1,6,15-trioxo -6,7-seco-ent-kaur-2-en-7, 20-olide.

Epi-eriocalyxin A (2) was obtained as a mixture which gave only one spot on TLC. The IR, UV and NMR spectra clearly showed that the mixture was composed of two quite similar ent-kaurane diterpenoids in a 1:1 ratio, and one of them was easily determined as eriocalyxin A (1). The comparison of 1 H and 13 C NMR signals of 2 with those of 1 indicated that 2 and 1 were epimeric at C-16. The β -orientation of Me-16 in 2 was confirmed by the upfield shift of C-12 at δ 17.6 and C-17 at δ 11.4 owing to the γ -steric compression effect between Me-16 β and H-12 β . Thus, compound 2 was identified as 16(R)-methyl-1,6,15-trioxo-6,7-seco-ent-kaur-2-en-7,20-oli de.

Maoccrystal N (3) contained three methyls, four methylenes (including one oxygen-bearing methylene), four methines, three quaternary carbons, two olefinic carbons and four carbonly carbons judging from its 1 H and 13 C NMR spectra. These facts, together with EI mass (ion at m / z 360 [M]⁺) exhibited 3 having a molecular formula of $C_{20}H_{24}O_6$.

The analysis of ¹H and ¹³C NMR data revealed that 3 resembled closely to eriocalyxin A (1) except for

A-ring. The carbon signal at δ 201.7 assigned to -CHO group in 1 was replaced by that at δ 172.4 arising from -COOH group. The proton signal at δ 10.08 corresponding to -CHO group in 1 was also disappeared in 3. All these facts showed the presence of -COOH in 3 instead of -CHO group in 1.

The results obtained from IR spectral data further confirmed the conclusion. The two absorption at 2860 and 2740 cm⁻¹owing to Fermi resonance of -CHO group in 1 was absent in 3, whereas a strong absorption at about 3000 cm⁻¹(br.) typical for hydroxly of carboxyl group was noticed in 3. Therefore, compound 3 was identified as 16(S)-methyl-6 α -carboxy-1, 15-dioxo-6, 7-seco-ent-kaur-2-en-7, 20-olide.

Maoecrystal O (4) was yielded as colorless crystals (from MeOH). Its 1 H and 13 C NMR spectral data were quite similar to those of maoecrystal L (5). The only difference between 4 and 5 was that the carbon signal at δ 203.8 arising from -CHO group in 5 was replaced by that at δ 173.9 from -COOH group in 4. Meanwhile, the proton signal at δ 9.98 corresponding to -CHO in 5 was absent in 4. The replacement of -CHO by -COOH group in 4 resulted in upfield shift of C-5 by 3.0 ppm.

This assignment was confirmed by the molecular ion peak at m/z 406 in EI mass spectra corresponding to $C_{22}H_{30}O_7$. The two absorption at 2850 and 2720 cm⁻¹characterized for the existence of CHO group in 5 was disappeared in 4 and there was a strong absorption at 3260 cm⁻¹owing to -COOH group being observed of the IR spectra in 4.

Table 1 ¹³C NMR data of maoecrystal L, eriocalyxin A*, laxiflorin C, and 1~4 in C_∗D_∗N (400 MHz, δ with reference to the signal of C_∗D_∗N)

Carbon	Maoecrystal L (5)	Eriocalyxin A*	1	Laxiflorin C	2	3#	4
2	24.3	125.1	125.0	124.5	125.1	124.6	24.7
3	39.8	156.6	157.5	158.9	157.5	157.2	39.5
4	34.1	36.2	36.2	36.7	36.2	35.9	34.0
5	61.6	57.5	57.5	47.4	57.6	54.3	58.6
6	203.8	200.1	201.7	58.2	201.7	172.4	173.9
7	170.0	169.1	169.6	175.5	169.7	169.6	170.2
8	59.3	59.8	59.9	59.5	59.2	59.1	59.4
9	43.6	42.6	42.6	41.8	42.8	43.1	45.2
10	43.7	50.0	50.6	52.3	50.4	50.2	43.1
11	17.5	17.8	18.1	20.2	20.0	18.0	17.8
12	29.4	29.7	30.0	17.9	17.6	29.9	29.8
13	35.5	35.4	35.7	32.6	32.6	35.4	35.6
14	28.8	29.5	30.0	32.7	32.8	29.4	28.8
15	216.1	216.2	216.7	217.5	217.0	216.2	216.5
16	50.9	51.1	51.0	48.8	48.9	51.0	51.2
17	16.0	16.8	16.7	11.8	11.4	16.6	16.7
18	32.6	31.5	31.2	31.7	31.1	31.9	33.5
19	23.8	24.3	24.0	23.8	24.1	24.7	23.3
20	68.0	68.7	69.3	70.8	69.5	69.3	68.8
OAc	170.0						170.2
	21.3						

[•] The data of eriocalyxin A reported by Wang et al., 1982

[#]The data were meausred in CDCl3 with reference to the signal of CDCl3

Accordingly, the structure of 4 was identified as 16(S)-methyl-1 α -acetoxy-6, α -carboxy, 15-ioxo-6,7-seco-ent-kauran-7, 20-olide.

EXPRIMENTAL

Mps: uncorr.. Optical rotations were taken on a JASCO-20C digital polarimeter. IR spectra were recorded with a Perkin-Elmer 577 spectrometer. UV spectra were obtained on a UV-210A spectrometer. MS spectra were measured on a VG Autospec spectrometer. NMR spectra were run on a Brucker AM- 400 spectrometer. The chemical shifts (δ) were expressed in ppm with reference to the solvent signals. Coupling constants (J) were given in Hz.

Plant Material Isodon eriocalyx (Dunn.) Hara was collected in Jiangchuan Prefecture, Yunnan Province, China, in September 10, 1994, and identified by Prof. H. W. Li. A voucher specimen was deposited in the Herbarium of Kunming Institute of Botany, Chinece Academy of Sciences, Kunming, P. R. China.

Extraction and Isolation Dried powdered leaves of *I. eriocalyx* (Dunn.) Hara (11.9 kg) were extracted with MeOH (4×401) under reflux and then concentrated in vacuum to give crude extract (978 g). The extract was suspended in H₂O and then successively fractioned with petroleum ether ($60 \sim 90^{\circ}$ C) and AcOEt. The AcOEt extract (395 g) was chromatographed on a silkica gel column (1.6 kg, $200 \sim 300$ mesh) eluting with CHCl₃by increasing amount of Me₂CO to yield seven fractions. Fraction I, II and III were submitted to MCI gel to decolor green pigments. Then each fraction was subjected to silkica gel column chromatography repeatedly followed by crystallization and finally afforded compounds: 1(3.1 g), 2(110 mg), 3(120 mg), 4(210 mg) and 5 (70 mg).

Eriocalyxin A (1), $C_{20}H_{24}O_5$, colorless crystals (MeOH), mp: 242~244°C; IR ν $^{\text{KBr}}_{\text{max}}$ cm $^{-1}$: 2956, 2860, 2740, 1736, 1721, 1705, 1664, 1300, 1211, 1089, 1060, 1045, 912 and 831; UV λ $^{\text{MeOH}}_{\text{max}}$ nm (log ε): 228.5 (4.03); EIMS (70 eV) m / z (%): 344[M]⁺(6), 316[M-CO]⁺(98), 301 (13), 287 (46), 269 (21), 257 (40), 227 (32), 165 (21), 1498 (43), 135 (100), 105 (41), 79 (65), 67 (46), and 55 (64); ^{1}H NMR (pyridine-d₅) δ: 5.94 (1H, d, J=10.0 Hz, H-2), 6.51 (1H, d, J=10.0 Hz, H-3), 10.08 (1H, d, J=3.2 Hz, H-6), 1.05 (3H, d, J=7.6 Hz, CH₃-16α), 1.27 (3H, s, CH₃-18), 1.16 (3H, s, CH₃-19), 4.98 (1H, d, J=11.2 Hz, H-20b) and 5.31 (1H, d, J=11.2 Hz, H-20a); ^{13}C NMR data see table 1.

Epi—**eriocalyxin A (2),** C₂₀H₂₄O₅, colorless crystals (MeOH); IR ν ^{KBr}_{max}cm⁻¹: 2910, 2840, 2740, 1730, 1703, 1660, 1450, 1360, 1245, 1208, 1085, 1040 and 910; UV λ ^{MeOH}_{max}nm: 228.5; EIMS (70 eV) m / z (%): 344 [M]⁺(4), 316 [M–CO]⁺(69), 301 (7), 287 (35), 269 (12), 257 (31), 277 (24), 164 (24), 135 (100), 107 (61), 91 (59), 79 (91), 67 (71) and 55(97); ¹H NMR (pyridine–d₅) δ: 5.94(1H, d, J = 10.0 Hz, H–2), 6.5(1H,d, J–10.0 Hz, H–3), 10.06(1H, d, J = 3.6 Hz, H–6), 1.00(1H, d, J = 7.2 Hz, CH₃–16β), 1.27(3H, s, CH₃–18), 1.16(3H, s, CH₃–19), 4.97(1H, d, J = 11.2 Hz, H–20b) and 5.32(1H, d, J = 11.2 Hz, H–20a); ¹³C NMR data see Table 1.

Maoecrystal N (3), $C_{20}H_{24}O_6$, Colorless crystals (MeOH), mp: 290~292°C, [α]_D²⁵+167.8° (C₅H₅N; c 0.359); IR ν $_{max}^{KBr}$ cm⁻¹: 3000 (br), 1750, 1740, 1700, 1400, 1310, 1270, 1240, 1212, 1160, 1110, 1060, and 930; UV λ_{max}^{MeOH} nm (log ε): 228.0 (3.93); EIMS (70 eV) m / z (%): 360 [M]⁺(3), 332 [M-CO]⁺(100), 301 (54), 79 (66), 67 (44) and 55 (75); HR FABMS (positive ion mode) m / z (%): 360.1532 (calc. 360.1573); ¹H NMR (CDCl₃) δ: 5.80(1H, d, J = 10.2 Hz, H-2), 6.56(1H, d, J = 10.2 Hz, H-3), 1.09(3H, d, J = 7.6 Hz, CH₃-16 α), 1.29(3H, s, CH₃-18), 1.27(3H, s, CH₃-19), 4.67(1H, d, J = 11.5 Hz, H-20b) and 5.11(1H, d, J = 11.5 Hz,

H-20a); ¹³C NMR data see Table 1.

Maoecrystal O (4), $C_{22}H_{30}O_7$, colorless crystals (MeOH), mp: $268 \sim 270^{\circ}$ C, $[\alpha]_D^{25} - 3.7^{\circ}$ (CHCl₃: c 0.410): IR ν_{max}^{KBr} cm⁻¹: 3260, 2940, 1760, 1728, 1686, 1360, 1223, 1072, 1032 and 910; EIMS (70 eV) m / z (%): 406 [M]⁺(3), 378 [M-CO]⁺(100), 362 (5), 349 (41), 318 (38), 290 (40), 272 (38), 164 (44), 149 (54), 135 (42), 120 (56), 105 (73), 83 (91), 67 (56) and 55 (87); HR FABMS (positive ion mode) m / z: 406.1992 (calc. 406.2028); ¹H NMR(pyridine-d₅) δ: 5.13(1H, d, J=3.3, 11.5 Hz, H-1 β), 0.98(3H, d, J=7.0 Hz, CH₃-16α), 1.24(3H, s, CH₃-18), 1.08(3H, s, CH₃-19), 5.39(1H, d, J=12.7 Hz, H-20b), 5.46(1H, d, J=12.7 Hz, H-20a) and 2.18(3H, s, OAc); ¹³C NMR data see Table 1.

Acknowledgments We are grateful to the analytical group of our laboratory for recording NMR, MS, UV, IR spectra and optical rotations.

REFERENCE

吴征镒,李锡文,宣淑洁等,1977. 中国植物志第 66 卷,唇形科(二). 北京:科学出版社,439~439 王宗玉,许云龙,1982. 毛萼香茶菜的新二萜成分. 云南植物研究,4:407~411

Hassan M G, Ghulam A M, 1994. Diterpenoids of Rabdosia species. Jour Chem Soc Pak, 16: 153~162

Shen X Y, Isogai A, Furihata K, Sun H D et al, 1994. 6, 7-Seco-ent-kaurane diterpenoid from Rabdosia eriocalyx.

Phytochemistry, 35: 820~821

Sun H D, Lin Z W, Niu F D, et al, 1995. Diterpenoids From Isodon eriocalyx var. laxi flora. Phytochemistry, 38: 1451~
1455

Takeda Y, Otsuka H, 1995. Recent advances in the chemistry of diterpenoids from Rabdosia species. Studies in natural Products Chemistry, 15: 111~184